On the Chemical Nature of Epicuticular Waxes in Some Succulent Kalanchoe and Senecio Species

Karsten Siemsa, Gerhard Jasa, F. Javier Arriaga-Giner^b, Eckhard Wollenweber^c and Marion Dörrc

- AnalytiCon Gesellschaft für Chemische Analytik und Consulting mbH, Gustav-Meyer-Allee 25, D-13355 Berlin, Bundesrepublik Deutschland
- Centro de Investigación y Desarrollo, Tabacalera S. A., E-28012 Madrid, Spain
- Institut für Botanik der Technischen Hochschule, Schnittspahnstrasse 3, D-64287 Darmstadt, Bundesrepublik Deutschland

Z. Naturforsch. 50 c, 451-454 (1995); received November 23, 1994/February 6, 1995

Kalanchoe spec., Senecio spec., Compositae, Leaf Wax, Pentacyclic Triterpenes

Leaves and other aerial parts of several succulent species each of Kalanchoe and Senecio exhibit more or less obvious wax coatings. The major components of these waxes were identified to be pentacyclic triterpenes. Some of these are rare natural products, while others are rather widespread.

Introduction

Most species of Kalanchoe (Crassulaceae) are more or less succulent xerophytes. The large Compositae genus Senecio also houses many species with succulent appearance. In both genera the presence of waxy material on their leaf surfaces and also on stem surfaces is rather widespread. In the course of our search for plants producing exudate flavonoids (c.f. Wollenweber, 1990) we have studied several species each of both genera. They all were found to be devoid of epicuticular flavonoid aglycones, but we happened to isolate some of the triterpenoid constituents forming the "leaf wax" (the term wax being used here in the botanical sense, regardless of the chemical definition; c.f. Barthlott and Wollenweber, 1981). Since to our knowledge externally accumulated triterpenoids have not been reported so far from neither Kalanchoe nor Senecio (Mahato et al., 1991), we studied the major constituents of their leaf waxes. As it turned out the wax does not only appear very sim-

Reprint requests to E. Wollenweber.

Telefax: (06151) 166878.

ilar in both genera, but also its composition is very similar, which suggests joint publication of these results.

Material and Methods

All plant material used in the present study was collected from plants cultivated in greenhouses of the Botanical Garden at Darmstadt. In each case fresh plant material was briefly rinsed with chloroform to dissolve the waxy epicuticular material. After evaporation of the solvent, the normally colourless residue was dissolved in toluene and applied to column chromatography on silica. Elution was done with toluene and increasing quantities of methylethyl ketone and methanol. Fractions were monitored by TLC on silica plates with solvents A (toluene-methylethyl ketone 9:1) and B (toluene-dioxane-glacial acetic acid 18:5:1). Terpenoids were visualized by spraying silica plates with MnCl₂ reagent, followed by heating (Jork et al., 1989). Mass spectra were recorded on a Varian MAT 311 at 70eV by direct inlet. ¹H-NMR, NOE, ¹³C-NMR and HMQC-spectra were recorded on a Bruker AC 400 (400 MHz respectively 100.6 MHz). All triterpenoids were identified by comparison of the ¹H-NMR data with those of authentic compounds or with literature data. MS-fragmentation patterns, especially Retro-Diels-Alder reactions, were also helpful for structure elucidation (cf. Budzikiewicz et al., 1963).

Results and Discussion

In most of the Kalanchoe and Senecio species analyzed here an epicuticular layer of waxy material is more or less apparent on leaves, on stems, and sometimes also on the bracts of the involucra. In Senecio kleinia Less, the wax is conspicuous on the succulent stems. In Kalanchoe thyrsiflora Harv. and particularly in K. pumila Baker the wax production is so rich that it forms a fragile chalky layer on the leaf surfaces. Long-chain saturated hydrocarbons were long known as components of this lipophilic material (Hegnauer, 1964), but the major wax constituents are obviously triterpenes. Those that we have identified are presented in Table IV and the results are briefly discussed in the following.

0939-5075/95/0500-0451 \$ 06.00 © 1995 Verlag der Zeitschrift für Naturforschung. All rights reserved.



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

452 Notes

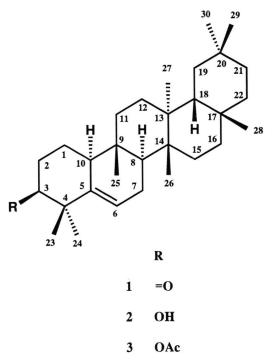


Fig. 1. Structural formulae of glutinone (1), glutin (2) and glutin acetate (3).

The leaf wax of *Kalanchoe fedtschenkoi* Hamet et Perr. de la Bâthie exhibits several terpenoid spots on TLC plates, with one dominating. The relevant product was isolated as colourless crystals. NMR studies showed it to be a mixture of ca 80% glut-5-en-3-β-ol (D:B-friedolean-5-en-3β-ol) with 20% friedelin. PMR signals of the former product are identical with those previously reported for this compound (Matsugana *et al.*, 1988). Friedelin is identified by comparison of its CNMR spectrum with that of authentic friedelin. Both triterpenes have been found earlier, along with taraxerol, in flowers of *Kalanchoe spathulata* (Gaind *et al.*, 1976), but their presence in the leaf wax had not been considered so far.

The wax of *Kalanchoe gastonis-bonnieri* Hamet et Perr. de la Bâthie exhibits one major terpenoid spot on TLC which is due to β -amyrenone, as the ${}^1\text{H-NMR}$ spectrum readily confirms. The same is true for *Kalanchoe thyrsiflora* Harv.

From the leaf wax of *Kalanchoe marnieriana* Jacobs. we isolated a mixture of two triterpenes, namely 90% of glutin (2) and 10% of friedelin.

Kalanchoe miniata. Hilsenb. et Boj. also exhibits a series of terpenoids, two of which have been crystallized. They were identified as β -amyrine acetate and glutinone (1).

From *Kalanchoe pumila* again a nonpolar triterpene was obtained in crystalline form. This was shown to be β -amyrenone. α -Amyrin, β -amyrin and sitosterol have been found as constituents of *Kalanchoe pinnata* long ago (Gaind and Gupta, 1972), but they were isolated from "pulverized leaves", so also in this case their external localization had not been noticed.

Table I. ¹H-NMR-data of glutinone (1), glutine (2) and glutinacetate (3) (CDCl₃, δ in ppm).

Proton	1	J [Hz]	2	J [Hz]	3	J [Hz]
1	1.84	dddd	*		*	
		(14; 4; 4; 4)				
1'	1.60	m	*		*	
2	2.38	ddd	*		*	
		(15; 4; 4)				
2'	2.46	ddd	*		*	
		(15; 10; 5)				
3	_		3.46	dd	4.69	dd
				(3; 3)		(3; 3)
6	5.69	m	5.64	m	5.56	m
7	1.97	m(2H)	*		*	
7 8	1.66	dd (12; 6)	*		*	
10	2.23	m	*		*	
18	1.59	m	*		*	
19	0.92	dd (13; 2)	*		*	
23	1.21	S	1.14	S	1.07	S
24	1.22	S	1.05	S	1.05	S
25	0.80	S	0.86	S	0.85	S
26	1.09	S	1.09	S	1.10	S
27	1.02	S	1.01	S	1.01	S
28	1.16	S	1.17	S	1.17	S
29	0.94	S	0.95	S	0.96	S
30	0.97	S	0.99	S	0.99	S
OAc	-	-	-	_	2.02	S

^{*} Overlapping signals.

Table II. Results of NOE-experiments on glutinone (1).

Irradiation in proton	NOE	
6	23	
7	25, 26	
8	27	
23	6	
24	10	
25	26, 1, 7	
26	28, 25, 18, 7	
27	8	
28	18, 26	
30	28	

Notes 453

Table III. 13 C-NMR of glutinone (1), glutine (2) and glutinacetate (3) (CDCl₃; δ in ppm, CDCl₃ = 77.0 ppm).

Carbon	1	2	3	
1	21.6	18.2	18.9	t
2 3	38.1	27.8	25.5	t
3	215.5	76.3	78.6	s/d
4 5	50.0	40.8	39.1	S
5	142.4	141.6	141.9	d
6	121.3	122.0	120.0	t
7	23.6	23.6	23.5	t
8	47.0	47.4	47.3	d
9	35.0	34.8	34.8	S
10	50.6	49.6	49.8	d
11	35.9	36.0	36.0	t
12	30.3	30.3	30.4	t
13	39.3	39.3	39.3	S
14	37.8	37.8	37.8	S
15	31.9	32.1	31.9	t
16	35.1	35.1	35.1	t
17	28.2	28.2	28.2	S
18	43.1	43.0	43.1	d
19	38.9	38.9	38.9	t
20	30.1	30.3	30.1	S
21	34.1	33.1	33.1	t
22	35.9	34.6	34.6	t
23	28.5	28.9	29.1	q
24	24.3	25.4	25.0	q
25	15.6	16.2	16.0	q
26	19.3	19.6	19.5	q
27	18.4	18.4	18.4	q
28	32.4	32.4	32.4	q
29	34.5	34.5	34.5	q
30	32.0	32.0	32.0	q
Ac	_	_	21.2	q
	_	-	170.9	s

The leaf wax composition of *Senecio cuneatus* Schultz Bip. and *Senecio ficoides* Schultz Bip. appeared identical. From various fractions we obtained three major crystalline materials, identified as friedelin, a mixture of ca 70% of glutin with 30% of taraxerol, and a mixture of ca 70% of taraxerol with 30% of taraxasterol. Taraxerol occurs rather rarely in Compositae, while taraxasterol is abundant in this family, but mostly known as a latex constituent (Hegnauer, 1964).

The wax layer of *Senecio kleinia* Less. exhibits a series of terpenoids. We obtained four crystalline

Table IV. Major triterpenoids in the leaf wax of Kalan-choe- and Senecio-species.

	β-Amyrenone	β-Amyrinacetate	3-Epilupeol	Friedelin	Glut-5-en-3 \beta-ol	Glutin (2)	Glutinacetate (3)	Glutinone (1)	Lupeone	Taraxerol	Taraxasterol	
Kal. fedtschenkoi				×	×							
Kal. gastonis-												
bonnieri	×											
Kal. marnieriana				×		\times						
Kal. miniata		\times						\times				
Kal. pumila	\times											
Kal. thyrsiflora	×											
Sen. cuneata				×		×				×	×	
Sen. ficoides				×		×				×	×	
Sen. kleinia			×	×			\times		×			

products which were identified as glutinyl acetate (3), friedelin, 3-epilupeol, and a mixture of some 60% of lupeone with 40% of friedelin.

Although glut-5-en-3β-ol, the corresponding acetate and the ketone glut-5-en-3-one are already known from different plant species, no assigned NMR-data have been reported so far. The ¹H-NMR data of glutinone (Table I) could partly be assigned by double resonance and NOE experiments (Table II). The conformation of glutinone in solution (determined by NOE) is the same as in solid state (determined by X-ray; Ohki *et al.*, 1981). The assignment of ¹³C-NMNR data (Table III) was done by 2D hetero correlated technics (HMQC) and comparison with those of friedelin derivatives (Patra and Chaudhuri, 1987; Prakasch *et al.*, 1987).

Acknowledgements

E. W. wishes to thank Mr. Klaus-Dieter Blümler and Mr. Helmut Groh from the Botanischer Garten der TH Darmstadt for their attention and for supplying plant material.

- Barthlott W. and Wollenweber E. (1981), Zur Feinstruktur, Chemie und Taxonomischen Signifikanz epicuticularer Wachse und ähnlicher Sekrete. Trop. Subtrop. Pflanzenwelt (Akad. Wiss. Lit. Mainz), Vol. 32, 35–97.
- Budzikiewcz H., Wilson J. M. and Djerassi C. (1963), Mass spectrometry in structural and stereochemical problems. XXXII. Pentacyclic triterpenes. J. Am. Chem. Soc. 85, 3688–3699.
- Hegnauer R. (1964), Chemotaxonomie der Pflanzen, Vol. 3. Birkhäuser Verlag Basel und Stuttgart.
- Jork H., Funk W., Fischer W. and Wimmer H. (1989), Dünnschichtchromatographie, Vol. 1a. Verlag Chemie, Weinheim.
- Gaind K. N., Singla A. K., Boar R. B. and Copsey D. B. (1976), Triterpenoids and sterols of *Kalanchoe spathulata*. Phytochemistry 15, 1999–2000.
- Gaind K. N. and Gupta R. L. (1972), Alkanes, alkanols, triterpenes and sterols of *Kalanchoe pinnata*. Phytochemistry 11, 1500–1502.

- Mahato S. B., Nandy A.K. and Roy G. (1992), Triter-penoids (Rev. Art. no. 67). Phytochemistry **31**, 2199–2249.
- Matsunaga S., Tanaka R. and Akagi M. (1988), Phytochemistry 27, 535-537.
- Ohki M., Tachibana K., Kuroda T., Takenaka A. and Sasada Y. (1981), Structure of alnusenone (D:B-friedoolean-5-en-3-one). Acta Cryst. **B** 37, 2092–2094
- Patra A. and Chaudhuri S. K. (1987), Assignment of carbon-13 nuclear magnetic resonance spectra of some friedelanes. Magn. Res. Chem. **25**, 95–100.
- Prakasch O., Roy R., Garg, H.S. and Bhakuni D. S. (1987), 13C NMR studies of the friedelane series of triterpenoids and the conformation of the D and E rings in friedelane-7-one. Magn. Res. Chem. 25, 39–41.
- Wollenweber E. (1990), On the distribution of exudate flavonoids among Angiosperms. Rev. Latinoamer. Quim. **21**, 115–121.